

## BTV TYPE 1 REAL TIME

Reference: ADI391-50 & ADI391-100

Test for the detection of Bluetongue Virus (BTV) of serotype 1 by real time enzymatic amplification

PCR Test – 50 & 100 reactions

For veterinary *in vitro* use only



Sample	Individual analysis	Pool of sample possible*, up to:
Blood	✓	✓
Organs	✓	✓

\*Depending on the epidemiological situation, sample quality and country-specific guidelines (refer to these guidelines).

## Kit composition

Content		ADI391-50 Kit	ADI391-100 Kit
		50 reactions	100 reactions
A5	Amplification solution	1 x 1000 µL tube with green cap (Ready to use)	2 x 1000 µL tubes with green cap (Ready to use)
BTV T1 CTL+	Bluetongue Virus serotype 1 positive control	1 tube with purple cap (To reconstitute)	1 tube with purple cap (To reconstitute)
NF-Water	Nuclease-Free Water	1 x 1000 µL tube with white cap (Ready to use)	1 x 1000 µL tube with white cap (Ready to use)

## Revision history

Date	Version	Modifications
01/2020	NE391-09	Addition of a "NF-Water" tube in the kit
11/2024	V01	New presentation. New reference with 100 reactions (ADI391-100) Addition of organ matrice Addition of "RNA FAST" program

Note: minor typographical, grammar and formatting changes are not included in the revision history.

## A. Introduction

The bluetongue virus is a non-contagious viral arthropod-borne infectious disease due to an Orbivirus (family Reoviridae, virus ARN), mainly transmitted by hematophagous midges from *Culicoides* genus.

The clinical expression is widely dependent on the environmental parameters (nutritional state, parasitism, and bacterial infections concomitant) and on the individual sensitivity. 35 distinct serotypes exist inducing partial or no cross protections between them. Transmission by pregnant ewes has also been described. Transmission by contaminated blood injection is possible when needles and syringes are re-used.

Samples for virus detection are bloods of animals with anticoagulants (EDTA). Virus is detected by isolation on embryonated eggs, *in vitro* cell culture, immunofluorescence on cell culture or by PCR.

## B. Test principle

ADIAVET™ BTV TYPE 1 REAL TIME test is based on the reverse transcription (RT) of RNA into complementary DNA. This reaction is followed by gene amplification of Bluetongue Virus serotype 1 specific DNA fragments. This test is intended to detect simultaneously, in one well:

- Bluetongue Virus serotype 1 (FAM labelled probe).
- GAPDH: internal control of extraction and amplification specific from an endogenous nucleic acid (HEX labelled probe or its equivalent).

This test is specific to the serotype 1. No cross-reaction has been observed with other types of BTV. No cross-reaction has been observed with EHDV strains.

## C. Storage conditions

On receipt, the kit should be stored at <-15 °C until the expiration date.

It is recommended to make fractions of A5 solution if it should be defrosted more than 3 times.

Do not thaw more than 3 times.

Store away from sunlight.

Do not mix reagents of two different batches.

## D. Material required but not provided

- Real-time Thermal cycler and device.
- Instrument for homogenous mixing of tubes.
- Pipettes of 1 - 10 µL, 20 - 200 µL and 200 - 1000 µL.
- Nuclease-free filtered pipette tips.
- Nuclease-free microtubes of 1,5 mL and 2 mL.
- Powdered-free latex or nitrile gloves.
- Nuclease-free water.
- Kit for nucleic acids extraction.

### Additional kits (on demand) for method adoption and PCR

- **LD<sub>PCR</sub> Positive Control – BTV type 1 (Ref.: ADC39LD)**  
Confirmation of performances – LOD<sub>PCR</sub> of kit.

## E. Warnings and precautions

- For veterinary *in vitro* use only.
- For animal use only.
- For professional use only.
- All instructions must be read before performing the test and strictly respected.
- Do not use reagents after the expiration date.
- Do not use reagents if the packaging is damaged.
- Do not open PCR wells or tubes after amplification.

- Do not mix reagents from different batches.
- Used material must be disposed of in compliance with the legislation in force regarding environmental protection and biological waste management.
- This kit contains products of animal origin. Certified knowledge of the origin and/or sanitary state of the animals does not totally guarantee the absence of transmissible pathogenic agents. It is therefore recommended that these products be treated as potentially infectious and handled observing the usual safety precautions (do not ingest or inhale).

## F. Nucleic acids extraction

### 1. Extraction kits

Nucleic acids must be extracted from the samples before using the kit. The RNA/DNA purification kits listed below are recommended by Bio-X Diagnostics:

Product name	Extraction system	Number of tests and reference
ADIAMAG™	Magnetic beads	200 tests: ref. NADI003 800 tests: ref. NADI003-XL

For the extraction, consult the user manual version, available on the website, indicated on the certificate of analysis included in the PCR kit of interest.

Extraction protocols are described in validation data. Other purification kits can be used if they have been validated by the user.

After extraction, nucleic acid extracts can be kept on ice or at +2/8 °C until use. For long term storage, they must be kept at a temperature below -15 °C or -65 °C.

### 2. Controls

Using controls allow to verify the reliability of the results. Controls can be included by series of analysis according to the recommendations defined by the standards in force (Cf. AFNOR U47-600...).

Controls	Validation of	How to proceed
No Template Control (NTC)	Absence of amplification contamination	5 µL NF-Water in a well per run
BTV T1 CTL+	BTV type 1 target amplification	5 µL CTL+ in a well per run
Negative extraction control	Absence of contamination for the extraction and amplification	1 extraction (water or lysis buffer) per run
Positive extraction control	Extraction and amplification	1 extraction (Positive sample between 1 et 100X LOD <sub>Method</sub> ) per run

## G. Procedure

### 1. Use of CTL+

- Add **200 µL** of « **NF-Water** » per tube.
- Homogenize the tubes by suction and pressure, then use a mixer, such as vortex, > 20 seconds and until dissolution of the blue pellet.
- After reconstitution, aliquot and store the solution at a temperature below -15 °C until the kit expiration date. Do not thaw more than 3 times.

For each assay, use **5 µL** of CTL+ denatured (see § « Denaturation ») in one of the dedicated wells (see § « Amplification », Step 2).

## 2. Denaturation of nucleic acids extracts

- For each sample and control(s), transfer minimum 10 µL of nucleic acids extracts in a tube or 96-plate and store the rest at a temperature below -15 °C or -65 °C.
- Incubate 3 minutes at +95 °C in a thermal cycler or heating block.
- Immediately transfer the tubes or 96-plate on melting ice or refrigerated block until use (to prevent RNA renaturation).
- To use, please refer to §« Amplification », Step 2.

## 3. Amplification

### Warning:

- Before starting, rehydrate or thaw reagents at room temperature in the dark.
- Homogenize all reagents and samples before use.
- Store reagents at a temperature below -15 °C after use.

**Step 1:** Dispense **20 µL** of amplification solution (A5) per well.

**Step 2:** Dispense **5 µL** of denatured nucleic acids extracts and **5 µL** of denatured controls in each dedicated well.

Use NF-Water for the No Template Control (NTC).

**Step 3:** Cover the wells with an appropriate optical film or caps.

**Step 4:** Start the PCR analysis.

The following programs are defined for ABI Prism thermocyclers (like 7500, QuantStudio5, Step-one...) from Applied Biosystems, for Mx3000, Mx3005P and AriaMx from Agilent, for LightCycler from Roche Diagnostics, for Rotor-Gene Q from Qiagen, for CFX96 and Chromo 4 from Biorad and for MIC from BioMolecular System.

RNA standard program		RNA FAST program	
10 min. 45 °C		10 min. 45 °C	
10 min. 95 °C		10 min. 95 °C	
15 sec. 95 °C*	40 cycles	5 sec. 95 °C	40 cycles
60 sec. 60 °C**		30 sec. 60 °C**	

\*30 sec. 95 °C for MX3000 and MX3005P.

\*\* Reading and parameters for fluorescence acquisition:

Fluorochrome	Absorbance (nm)	Emission (nm)
FAM	494	520
HEX or equivalent	530	549
ROX	575	602

**Note:** The Quencher is non-fluorescent. The A5 solution contains a passive reference read in the same spectra as ROX for ABI machines.

For other thermal cycler instruments, please contact your sales representative or the customer relations department.

## H. Interpretation of results

Display all curves and position the threshold line for each fluorochrome.

### 1. Test validation

Amplification is valid if the following results are obtained.

Expected Ct (Threshold Cycle) values for the CTL+ are indicated on the certificate of analysis of the kit.

Controls	Amplification		Validation of
	FAM	HEX or equivalent	
No Template Control (NTC)	No	No	Absence of amplification contamination
BTV T1 CTL+	Yes	Yes/No	Target amplification
Extraction negative control	No	No	Absence of extraction contamination
Extraction positive control	Yes	Yes	Extraction and amplification steps

## 2. Results interpretation

Nucleic acids extraction and amplification are **valid** for each sample if at least one typical amplification curve is observed in FAM and/or HEX or equivalent.

Amplification		Interpretation
FAM	HEX or equivalent	BTV TYPE 3
No	Yes	Not detected
Yes, Ct < 34	Yes	Detected
Yes, 34 ≤ Ct < 40	Yes	Detected in small quantities
No	No/yes	Not determined

**“Detected in small quantities”:** The infection status cannot be defined: very recent infection (start of viraemia) or old infection (end of viraemia).

**“Not determined”:** The results cannot be interpreted for the corresponding sample.

**Possible causes:**

Defective PCR due to inhibitors, set-up error, absence of samples, degraded samples and/or

Issue with nucleic acid extraction (loss or destruction of nucleic acids).

**Recommendations:**

Set up a new PCR assay using pure nucleic acid extracts and dilute to a 1/5 concentration in nuclease-free water;

If the test is still invalid, repeat the nucleic acid extraction by diluting the blood by half in PBS buffer. Finally, if the result cannot be interpreted, the sample cannot be used (presence of RT-PCR inhibitors; lysed or putrefied sample, etc.).

**Note: This RT-PCR test does not exclude the presence of other BTV serotypes than serotype 1 in the analysed sample.**

## Symbols

Symbols	Signification
	Catalog number
	Manufacturer
	Temperature limitation
	Use by
	Batch code
	Consult Instructions for Use
	Contain sufficient for "n" tests
	For veterinary <i>in vitro</i> use only – For animal use only
	Keep away from sunlight

1 | Extract nucleic acids with

**Adia<sup>X</sup>  
Mag**

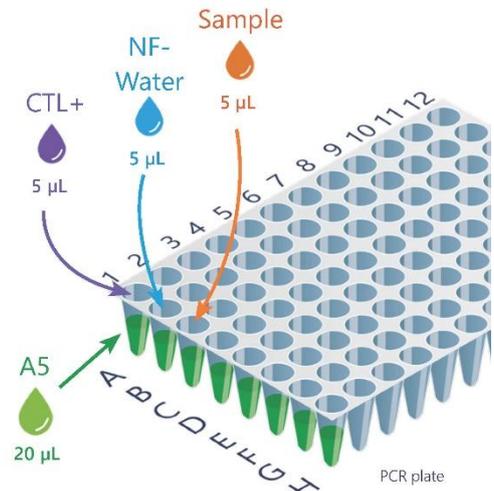


Scan me to discover Adiamag™

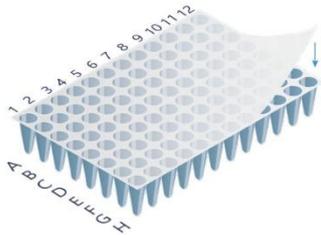
2 | Distribute **20 µL** of **A5** amplification solution

3 | Distribute **5 µL** of **nucleic acids**, **CTL+** and **NF-Water**

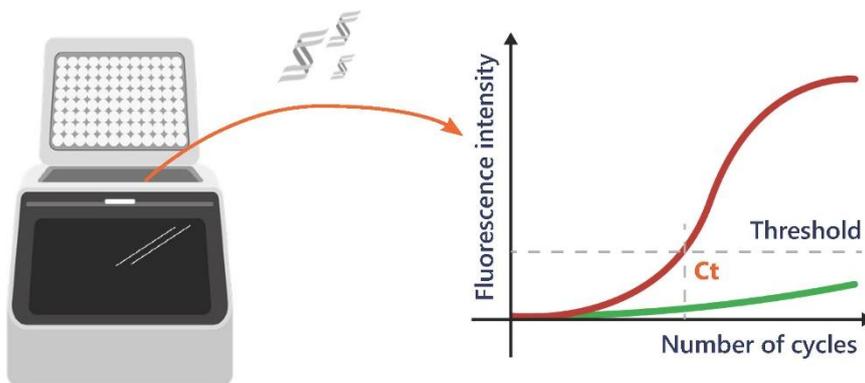
*\*Previously denatured 3 min. 95°C*



4 | Seal the wells



5 | Start PCR analysis



*\*The notes do not replace the instructions for use of which they are a summary.*